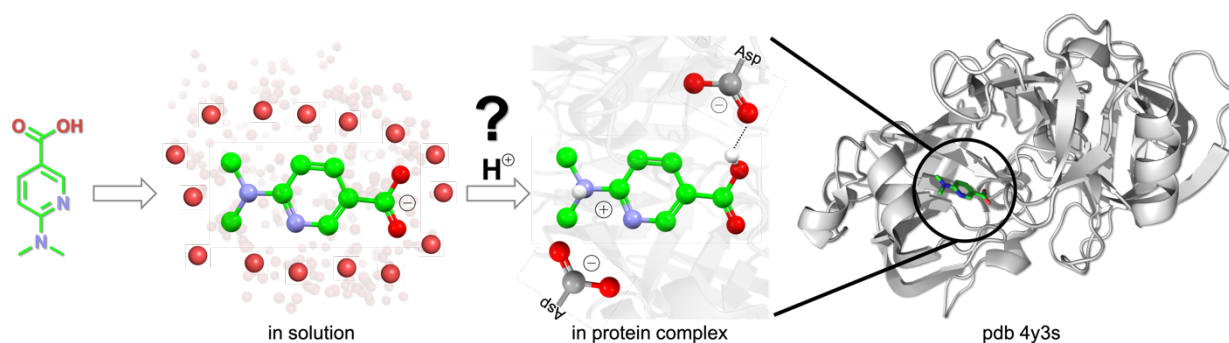


# Protonation effects of endothiapepsin-fragment complexes

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Endothiapepsin (EP) is an aspartic protease isolated from the ascomycete *Cryphonectria (Endothia) parasitica* and is often used as a milk clotting enzyme in cheesemaking processes. [1] Furthermore, EP often serves as a surrogate for renin (blood pressure) and  $\beta$ -secretase (Alzheimer's disease) in structure-based inhibitor design. [2] Due to a straightforward purification of EP with high yields, we use it as a model enzyme for the investigation of protonation changes upon protein-ligand complex formation (see figure below). Protonation changes upon binding of a ligand to a protein is often an overlooked phenomenon of molecular recognition. In fact, changes in protonation of titratable groups of either the ligand or the protein can have a serious impact on the binding geometry as well as the binding affinity. [3] In order to experimentally determine protonation changes, we perform Isothermal Titration Calorimetry (ITC) experiments with known EP-binding fragments in various buffers with differing heats of ionization. By supporting the ITC measurements with  $pK_a$  calculations (Poisson-Boltzmann calculations) we can predict the residue or ligand functional group responsible for the protonation change. [3] In a next step, we additionally want to determine the binding mode of the fragments via X-ray crystallography.



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